Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

CHROMIUM (III) COMPLEXES AND THEIR RELATIONSHIP TO THE GLUCOSE TOLERANCE FACTOR

A THESIS PRESENTED IN PARTIAL FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF • MASTER OF SCIENCE IN CHEMISTRY AT MASSEY UNIVERSITY NEW ZEALAND

JUAN ANTON COOPER

1982

83.04685

664.03 MASSEY UNIVERSITY LIBRARY 63.04685

ABSTRACT

To forms of the dinicotinate complex $Cr(nic)_2(H_2O)_3OH$ were formed which were yellow and blue, respectively. For the yellow form the nicotinic acid ligands were coordinated via the pyridine ring nitrogen atom but this complex was biologically inactive, while for the blue form nicotinic acid was coordinated via the carboxylate group and this compound was biologically active. Only Cr(III) formed a stable carboxylate coordinated dinicotinate complex. No stable complexes were formed with Fe(III) and Mn(III) due to significant olation, even at acidic pH's, and the complexes of nicotinic acid with Cr(II), Mn(II), Co(II) and Ni(II) were all pyridine nitrogen atom coordinated and biologically inactive.

Several chromium (III) complexes with amino acids possessed biological activity also, and these included the α -carboxylate coordinated species $Cr(gly)_n (H_2 0)_{6-n}^{3+}$ and $Cr(glu)_n (H_2 0)_{6-n}^{3+}$, the bidentate coordinated $Cr(gln)_2$ $(H_2 0)_2^+$ complex, and the NH₄OH- eluted complexes obtained when $Cr(gly)_2(H_2 0)_2^+$ and $Cr(glu)_2(H_2 0)_2^+$, but not $Cr(cys)_2$ $(H_2 0)_2^+$, were eluted from a DOWEX 50W-X12 cation-exchange column (loss of the α -amino coordination was postulated to have occurred).

The biologically active mixed ligand complex postulated as $Cr_2(nic)_4(gly)_2(OH)_2$ was prepared and found to be stable at neutral pH as a result of coordination of the glycine ligands.

The activity of the chromium (III) complexes in the yeast fermentation assay suggested that similar effects would be found in mammalian systems. The yeast assay system was found to be a simple, quick and reproducible method of determining biological activity.

All of the active chromium (III) complexes prepared were found to be similar, in structure, to the diguanide compound 1,4-diguanidinobutane which is known to lower blood sugar levels in mammals. This similarity in structure suggested a similar function might be possessed by the complexes reported in this thesis.

ACKNOWLEDGEMENTS

I wish to thank my supervisors Dr Len F. Blackwell and Dr Paul D. Buckley for their invaluable advice and assistance and the encouragement offered throughout the course of this work.

Thanks are also extended to all members of the Chemistry, Biochemistry and Biophysics Department for their help especially: Dr Steven J. Haylock, Mrs Rose Motion, Dr Alastair K.H. MacGibbon and Dr Eric W. Ainscough for their assistance and advice.

Thanks also to friends and family for the support and encouragement received during this study, especially my wife, Judith.

Finally, I would like to thank Mrs Veronica Fieldsend for the excellent job in typing this thesis.

TABLE OF CONTENTS

ii iv ACKNOWLEDGEMENTS

TABLE OF CONTENTS	v
LIST OF FIGURES	

LIST OF TABLES

ABSTRACT

SECTION ONE INTRODUCTION

		SECTION TWO	5
	1	NICOTINIC ACID COMPLEXES WITH CHROMIUM	
		AND OTHER TRANSITION METALS	
2.1	Introd	uction	5
2.2	Transi	· tion Metal Chemistry	6
			6
	ALL CONTRACTOR OF A	Magnetic Properties	7
	2.2.3	Electronic Structures and Spectral Properties	8
	2.2.4	Charge Transfer Spectra	15
	2.2.5	Electron Spin Resonance Spectra	15
2.3	Method	s and Materials	16
	2.3.1	Sources of Chemicals	16
	2.3.2	Methods and Instrumentation	16
		2.3.2.1 Micro analysis	16
		2.3.2.2 Chromium determination	16
		2.3.2.3 Electronic, infra-red and electron spin resonance spectroscopy	17
		2.3.2.4 Magnetic susceptibility	17
		2.3.2.5 X-ray diffractometry	17
		2.3.2.6 Absorbance, pH and conductivity measurement	17
		2.3.2.7 Ion-exchange resins	17

Page

vi

		2.3.2.8	Gel filtration resin	18
2.4	Results	5		18
	2.4.1	The Yell Complex	ow Chromium (III) Dinicotinate	18
	4	2.4.1.1	Preparation of yellow chromium (III) dinicotinate complex	18
		2.4.1.2	X-ray powder diffractometry	21
		2.4.1.3	Electronic spectra and magnetic properties	21
		2.4.1.4	Electron spin resonance spectroscopy	25
	8	2.4.1.5	Infra-red spectra	25
	2.4.2	Other Tr Complex	cansition Metal Nicotinic Acid	28
		2.4.2.1	Preparation of Co(nic) ₂ (H ₂ O) ₄	28
		2.4.2.2	Preparation of Ni(nic) ₂ (H ₂ O) ₄	28
		2.4.2.3	Preparation of Mn(nic) ₂ (H ₂ O) ₄	28
		2.4.2.4	Preparation of Fe(nic) (H ₂ O) OH	29
			Properties of the transition metal complexes	29
		2.4.2.6	Electronic and magnetic properties	30
		2.4.2.7	Infra-red spectra .	31
		2.4.2.8	X-ray diffraction powder patterns	31
	2.4.3	The Blue	e Chromium Dinicotinate Complex	35
			Preparation of blue chromium dinicotinate complex	35
		2.4.3.2	Electronic spectra and magnetic properties	35
		2.4.3.3	Infra-red spectra	36
		2.4.3.4	Electron spin resonance spectroscopy	39
	×	2.4.3.5	X-ray powder diffractometry	39
	2.4.4		n of the Blue Chromium-Nicotinic Acid x with Acid	40
		2.4.4.1	Electronic spectra	40
		2.4.4.2	Ion-exchange and gel filtration chromatography	41
	2.4.5	The Solution ate Con	uble Blue Chromium (III)-Mononicotin- mplex	41
		2.4.5.1	Attempted preparation of a mono- nicotinate complex of chromium (III) 4 1

		2.4.5.2 Electronic spectra	46
2.5	5 Discussion		
	2.5.1	Yellow Chromium (III)-Dinicotinate Complex 2.5.1.1 Comparison with literature	46 53
	2.5.2	Other Transition Metal Complexes with Nicotinic Acid	57
	2.5.3	Blue Coloured Chromium (III)-Dinicotinate Complex	60
	2.5.4	The Reaction of the Blue Cr(nic) ₂ (H ₂ O) ₃ OH Complex with Acid	65
	2.5.5	The Soluble Blue Mononicotinate Complex of Chromium (III)	69
	2.5.6	Comparisons with Literature	75
2.6	Conclu	sion	78

SECTION	THREE	82

AMINO ACID COMPLEXES OF CHROMIUM (III)

3.1	Introd	luction	82
3.2	Method	ls and Materials	83
	3.2.1	Sources of Chemicals	83
	3.2.2	Experimental Methods and Instrumentation	83
		3.2.2.1 Cation-exchange chromatography	83
		3.2.2.2 Anion-exchange chromatography	84
		3.2.2.3 Gel filtration chromatography	84
		3.2.2.4 Electronic spectroscopy	84
		3.2.2.5 Absorbance, pH and conductivity measurement	84
		3.2.2.6 Chromium determination	85
		3.2.2.7 Micro analysis	85
3.3	Result	S	85
	3.3.1	Chromium (III) Complexes with Glycine	85

		3.3.1.1	Preparation of glycine complexes	85
		3.3.1.2	Ion-exchange chromatography of glycine complexes	86
		3.3.1.3	Electronic Spectra	88
	3.3.2	Chromium	(III) Complexes with Cysteine	89
		3.3.2.1	Preparation of cysteine complexes	89
		3.3.2.2	Ion exchange chromatography of cysteine complexes	90
		3.3.2.3	Electronic spectra of cysteine complexes	91
		3.3.2.4	Titration of NaCr(cys) ₂ .2H ₂ O	93
		3.3.2.5	Determination of cysteine by the DTNB reaction	98
	3.3.3	Chromiun	(III) Complexes with Glutamic Acid	100
		3.3.3.1	Preparation of glutamic acid complexes	100
		3.3.3.2	Ion exchange of glutamic acid complexes	100
		3.3.3.3	Electronic spectra	103
	3.3.4	Chromium	n (III) Complexes with Glutamine	104
		3.3.4.1	Preparation of $[Cr(NH_3)_6]$ (NO ₃) ₃	104
		3.3.4.2	Preparation of glutamine complexes	105
		3.3.4.3	Ion exchange of the glutamine complexes	106
		3.3.4.4	Electronic spectra	106
3.4	Discuss	sion		107
	2 1 1	Chromiur	(III) Complexes with Clusing	107
	3.4.1		n (III) Complexes with Glycine The tris-glycine chromium (III)	107
			complex	107
		3.4.1.2	The dimer bis-glycine chromium (III) complex	109
		3.4.1.3	Monodentate glycine complexes with chromium (III)	111
		3.4.1.4	The soluble bis-glycine chromium (III) complex	114
		3.4.1.5	The reaction of Cr(gly) ₂ (H ₂ O) ₂ ⁺ with NH ₄ OH	n 118

	3.4.2	Chromium	(III) Complexes with Cysteine	119
		3.4.2.1	The tridentate cysteine complex with chromium (III)	119
		3.4.2.2	The reaction of NaCr(cys) ₂ .2H ₂ O with acid	124
	91	3.4.2.3	The reaction of NaCr(cys) ₂ . ^{2H} 2 ^O with base	128
		3.4.2.5	The pH dependent equilibrium between Cr(cys) ₂ (H ₂ O) ₂ and Cr(cys) ₂	129
	3.4.3	Glutamic	Acid Complexes with Chromium (III)	130
		3.4.3.1	The soluble pink glutamic acid complex with chromium (III)	130
		3.4.3.2	The soluble purple glutamic acid complex with chromium (III)	133
		3.4.3.3	The soluble blue monodentate glutamic acid complex with chromium (III)	135
		3.4.3.4	The effect of ammonia on the cationic glutamic acid complexes of chromium (JII)	136
	3.4.4	Glutamir	e Complex with Chromium (III)	136
		3.4.4.1	The soluble red glutamine complex with chromium (III)	136
		3.4.4.2	The soluble purple glutamine complex with chromium (III)	139
3.5	Conclu	sion	×	142

7 N.

SECTION FOUR 146

MIXED LIGAND COMPLEXES WITH CHROMIUM (III)

4.1	Introd	Introduction		
4.2	Method	s and Mat	erials	148
	4.2.1	Sources	of Chemicals	148
	4.2.2	Methods	and Instrumentation	148
		4.2.2.1	Chromium determination	148
		4.2.2.2	Electronic spectroscopy	148
		4.2.2.3	Absorbance, pH and conductivity measurement	148

4.2.2.4	Ion-exchange resins	148
4.2.2.5	Gel filtration resins	148

4.3	Results	5	149
	4.3.1	Preparation of a Mixed Solution of Chromium (III) Complexes with Nicotinic Acid and Glycine	149
		4.3.1.1 Ion-exchange chromatography	149
		4.3.1.2 Gel filtration chromatography	150
		4.3.1.3 Electronic spectra	150
	4.3.2	Preparation of a Mixed Solution of Chromium (III) Complexes with Nicotinic Acid and Cysteine	150
		4.3.2.1 Ion-exchange chromatography	152
		4.3.2.2 Gel filtration chromatography	152
		4.3.2.3 Electronic spectra	152
	4.3.3	Preparation of Chromium (III)-glutathione	150
		Complexes	153
		4.3.3.1 Ion exchange chromatography and electronic spectra	153
	4.3.4	Preparation of a Mixed Solution of Chromium (III) Complexes with Nicotinic Acid and Reduced Glutathione	153
		4.3.4.1 Gel filtration chromatography and electronic spectra	154
	4.3.5	Preparation of a Mixed Solution of Chromium (III) Complexes with Nicotinic Acid, Glycin Cysteine and Glutamic Acid	ne, 154
		4.3.5.1 Ion-exchange separation	155
		4.3.5.2 Gel filtration chromatography	157
		4.3.5.3 Electronic spectra	158
		4.3.5.4 Thin layer chromatography	161
4.4	Discus	sion	164
	4.4.1	Mixed Ligand Complexes of Chromium (III) with Nicotinic Acid and Glycine	164

4.4.2 Mixed Ligand Complexes of Chromium (III) with Nicotinic acid and Cysteine 168
4.4.3 Chromium (III) Complexes with Glutathione 171

х

	4.4.4	Mixed Ligand Complexes of Chromium (III) with Nicotinic Acid and Reduced Glutathione	173
	4.4.5	Mixed Ligand Complexes of Chromium (III) with Nicotinic Acid, Glycine, Cysteine, and Glutamic Acid	174
4.5	Conclu	sion	177
		SECTION FIVE	180
	THE BIC	DLOGICAL ACTIVITY OF CHROMIUM (III) COMPLEXES	
5.1	Introd	uction	180
5.2	Method	s and Materials	184
	5.2.1	Yeast culture	184
	5.2.2	Pre-assay growth on plating medium	184
	5.2.3	Pre-assay growth in a liquid medium	184
	5.2.4	Assay growth in a defined medium	185
	5.2.5	Cell harvesting and concentration determin- ation	185
	5.2.6	Standard assay technique	186
	5.2.7	Manometric calculations and data inter- pretation	186
	5.2.8	Optimum assay conditions	188
5.3	Result	S	189
	5.3.1	Nicotinic Acid Complexes with Transition Metals	189
	5.3.2	Chromium (III) Complexes with Various Amino Acids	192
	5.3.3	Mixed Ligand Complexes of Chromium (III)	193
	5.3.4	The Effect of Ammonium Hydroxide on Chromium (III) Complexes	195
5.4	Discus	sion	199
	5.4.1	Nicotinic Acid Complexes with Transition Metals	199
	5.4.2	Chromium (III) Complexes with Various Amino Acids	200

5.4.3	Mixed	Ligand	Complexes	with	Chromium	(III)	201
-------	-------	--------	-----------	------	----------	-------	-----

5.4.4 The Effect of Ammonium Hydroxide on Chromium (III) Complexes 202

SECTION SIX 205 CONCLUSION

APPENDIX	LIST	OF	ABBREVIATIONS	211	

REFERENCES

212

LIST OF FIGURES

Figure	Page
1.1 Hypothetical ternary complex of chromium at site of action	3
2.1 Energy level diagram of an octahedral metal	ion 11
2.2 Ground state d-orbital configurations	11
2.3 Tanabe-Sugano diagrams	13
2.4 Apparatus for preparation and reaction of C	r ²⁺ 19
2.5 Infra-red absorption spectra of (a) yellow $Cr(nic)_2(H_2O)_3OH$, (b) $Co(nic)_2(H_2O)_4$, and (c) nicotinic acid	20
2.6(a) Electron spin resonance spectrum of yellow Cr(nic) ₂ (H ₂ O) ₃ OH	26
2.6(b) Electron spin resonance spectrum of blue Cr(nic) ₂ (H ₂ O) ₃ OH	26
2.7 Infra-red absorption spectrum of (a) $Mn(nic (H_2O)_4, (b) nicotinic acid, (c) Ni(nic)_2 (H_2O)_4, (d) Co(nic)_2(H_2O)_4, and (e) yellow Cr(nic)_2(H_2O)_3OH$) ₂ 32
2.8 Infra-red absorption spectrum of (a) blue Cr(nic) ₂ (H ₂ O) ₃ OH and (b) nicotinic acid	37
2.9 Elution profile of a blue coloured $Cr(nic)_2$ (H ₂ O) ₄ ³⁺ complex on a Sephadex G15 column	42
2.10 Elution profile of the blue coloured chromium (III)-nicotinic acid species on a Sephadex G15 column	45

Figure	Page
2.11 Molecular structure of Co(II)(nic) ₂	(H ₂ O) ₄ 47
2.12 Structure of yellow Cr(nic) ₂ (H ₂ O) ₃ O	Н 48
2.13 Proposed structures of dinicotinate with bridging halogens	complex 55
2.14 Structure of the blue $Cr(nic)_2(H_2^0)$	3 ^{OH} complex 66
2.15 Structure of the blue coloured Cr(n species in acidic solution	ic) ₂ (H ₂ O) ₄ ³⁺ 68
2.16 Structure of soluble blue Cr(nic)(H	2 ⁰) ₅ ²⁺ 70
2.17 Reaction scheme for dimeric chromius nicotinic acid complex	m (III)- 72
2.18 Predicted overlap of the elution protected blue coloured $Cr(nic)_2(H_2O)_4^{3+}$ $(H_2O)_5^{2+}$ species on a Sephadex G15	and Cr(nic)
3.1 Electronic spectrum of NaCr(cys) ₂ .2	H ₂ O 92
3.2(a) Series of electronic spectra obtain aqueous solution of NaCr(cys) ₂ .2H ₂ O was titrated with dilute acid	
3.2(b) Series of electronic spectra obtai an aqueous solution of NaCr(cys) ₂ .2 was titrated with dilute base	
3.3 Variation in intensity of the d-d b visible spectrum of an aqueous solu NaCr(cys) ₂ . ^{2H} 2 ^O with pH	
3.4(a) Titration curve for an aqueous sol NaCr(cys) ₂ . ^{2H} 2 ^O	ution of 97

Figure	Page
3.4(b) Titration curve for an aqueous solution of cysteine HCl	97
3.5 Plot of absorbance at 412nm (for the DTNB anic against time for the reaction of NaCr(cys) ₂ . ^{2H} ₂ at pH 8.0 with DTNB	
3.6 Guggenheim plot of ln(A-A') against time derived from the absorbance plot (Figure 3.5)	101
3.7 Structure of Cr(gly) ₃ .H ₂ O	109
3.8 Structure of [Cr(gly)2 ^{OH]} 2	111
3.9 Structure of postulated Cr(gly) ₂ (H ₂ O) ₄ ³⁺	113
3.10 Structure of Cr(gly) ₂ (H ₂ O) ₂ ⁺	116
3.11 Structure of NaCr(cys) ₂ . ^{2H} 2 ^O	120
3.12 Structure of red coloured Cr(cys)2(H20)2+	127
3.13 Structure of Cr(glu) ₂ (H ₂ O) ₂ at pH 8.5	132
3.14 Reaction of chromium (III) with glutamic acid under acidic conditions	134
3.15 Structure of Cr(gln) ₂ (H ₂ O) ₂ ⁺	138
3.16 Structure of Cr(pyr) ₂ (H ₂ O) ₄ ⁺	141
4.1(a) Elution profile of the chromium (III)- nicotinate-glycine reaction mixture on a Sephadex G15 column	151
4.1(b) Elution profile of the dicysteine chromium (III)-nicotinic acid reaction mixture on a	

Sephadex G15 column

151

xv

Page

Figure

4.1(c)	Elution profile of the chromium (III)- nicotinate-glutathione reaction mixture on a Sephadex G15 column	151
4.2(a)	Elution profile of the cationic chromium (III) complexes prepared in Section 4.3.5 on a DOWEX 50-X12 column	156
4.2(b)	Elution profile of the cationic chromium (III) complexes prepared in Section 4.3.5 on a DOWEX 50-X12 column	156
4.3(a)	G10 column eluted with water	159
4.3(b)	G10 column eluted with 50% EtOH	159
4.3(c)) Elution profile of fraction G ₁ on a Sephadex G10 column eluted with water	159
4.4	pH titration of reaction mixture	160
4.5	Summary of the cation-exchange chromatography of the purple $Cr(nic)_2(H_2O)_4^{3+}$ -glycine reaction mixture	164
4.6	Postulated polymeric structure of [Cr(nic) ₂ (OH) ₂] _n	166
4.7	Postulated structure of $Cr_2(nic)_4(gly)_2(OH)_2$	167
4.8	Summary of the cation-exchange chromatography of the purple $Cr(cys)_2(H_2O)_2^+$ -nicotinic acid reaction mixture	168

1201020-001		
V 17		
~ V	-	-
	_	_

Figure		Page
4.9	Postulated structure of Cr(cys) ₂ (nic) ₂	170
4.10	Structure of Cr(glut) ₂ (H ₂ O) ₂	172
5.1	Standard curves used in the determination of yeast cell concentration	187
5.2	Standard yeast assay showing carbon dioxide production	190
5.3	Activity of the NH4OH-eluted Cr(gly)2(H2O)2 ⁺ species as a function of chromium (III) concentration	197
5.4	The structure of active monodentate chromium (III)-amino acid complexes	204
6.1	Comparison of the structures of the active chromium (III) complexes	209

F

LIST OF TABLES

Table		Page
2.1	Electronic configuration of metal atoms	7
2.2	"Spin only" magnetic moments for various numbers of unpaired electrons	8
2.3	Theoretical and experimental magnetic moments for various transition metal ions	9
2.4	Splitting of Russell-Saunders states	12
2.5	Analytical data of nicotinic acid complexes	20
2.6	X-ray diffraction powder patterns of yellow $Cr(nic)_2(H_2O)_3OH$ and $Co(nic)_2(H_2O)_4$	23
2.7	Electronic absorption bands of nicotinic acid complexes	24
2.8	Magnetic properties of nicotinic acid complexes	24
2.9	Infra-red spectrum of the yellow chromium- nicotinic acid complex	27
2.10	Infra-red spectra of nitrogen coordinated dinicotinic acid complexes	33
2.11	X-ray diffraction powder patterns of Ni(nic) ₂ $(H_2O)_4$ and Mn(nic) ₂ $(H_2O)_4$	34
2.12	Infra-red spectrum of blue chromium-nicotinic acid complex	38
3.1	Analytical data for amino acid complexes	85

Π-	h	1	-
Τa	10	Te	2

Page

3.2	Electronic spectra of glycine complexes	89
3.3	Electronic spectra of cysteine complexes	91
3.4	Reaction conditions for preparation of glycine complexes	115
3.5	Summary of amino acid complexes with Chromium (III)	143
4.1	Cationic fractions eluted from DOWEX 50-X12 column	157
4.2	Electronic spectra of cationic fractions	161
4.3	Thin layer chromatography of amino acids and nicotinic acid	162
4.4	Thin layer chromatography of non-chromatographed P fraction	163
4.5	Thin layer chromatography of G_1 fraction	163
5.1	Activity of nicotinic acid complexes of transition metals	191
5.2	Activity of chromium (III) complexes with various amino acids	192
5.3	Activity of amino acids	193
5.4(a)	Activity of mixed ligand complexes of chromium (III)	194
5.4(b)	Activity of mixed ligand complexes of chromium (III)	194

5.5	Activity of Chromium	(III)	complexes	after	
	elution with NH_4OH				196
5.6	Activity of Chromium	(III)	complexes	after	
	elution with NH ₄ OH				198

.

xx