Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author. THE FIBRILLAR ORGANIZATION OF COLLAGEN

IN CONNECTIVE TISSUE

A thesis presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Biophysics at Massey University

ALAN SMITHSON CRAIG

1984

80.101

DEDICATION

This thesis is dedicated to my wife - Wendy and our children Michael, Kim, Kirsten and Hadley.

.

ABSTRACT

Although certain aspects of connective tissue structure have been studied in considerable detail, comparatively little effort has been devoted to studying one of the largest structural units present in tissues - the collagen fibril. In this thesis electron most microscope observations have been made on the transverse dimensions of fibrils from tissues as diverse as cornea, skin and tendon. Collagen fibril diameter distributions have been measured for such tissues from a wide range of animals - predominantly mammals, but also fish, amphibians, reptiles and birds - at varying stages of development. These data have allowed the growth of collagen fibrils to be studied quantitatively and their size distributions to be related to their mechanical attributes. Diseased tissues or tissues containing anomalous fibril diameter distributions have also been studied and, where possible, the data have been related to the altered mechanical properties of the tissue and to its mode of growth and development. In a coordinated study with other research workers, the content of the individual glycosaminoglycans in a tissue have been shown to be related to the mass-average diameters of the collagen fibrils in those tissues. These results provide a basis for understanding the feedback mechanism by which fibril size distributions may be modified in line with changing mechanical needs and indicate the fundamental steps in the growth and development of fibrils.

In addition to these studies, two other specific problems were addressed. In the first, the ultrastructure of a specialized connective tissue – the cornea – was studied in detail. By maintaining precise experimental protocols and measurement procedures it was shown, contrary to the previous data of others, that the

mammals, birds, reptiles, amphibians and collagen fibrils in cartilaginous fish were similar to one another but significantly different to the corneal stromal fibrils of the bony fish. Further studies, which indicated that the fibrils were constant in diameter across the width of the stroma, clarified previous results which had indicated a gradual change in diameter with varying depth in the stroma. An age-related study of fibril diameters in the cornea was also undertaken. The second problem investigated was the degree of shrinkage introduced during the preparative procedures for electron microscopy. In collaborative studies with others, X-ray and electron microscope observations were made on the same tissue in hydrated and dehydrated states respectively. Analyses of these data indicated that significant lateral shrinkage does indeed occur in fibrils from foetal or immature tissues as well as in mature tissues containing only small diameter fibrils. Throughout the thesis possible sources of artefact introduced by the technique of electron microscopy have been considered and the data interpreted conservatively.

ACKNOWLEDGEMENTS

Many people have helped me in a variety of ways to get this thesis into its present form.

In my earlier years at DSIR Keith Williamson introduced me to, and gave me sound guidance in, the principles and techniques of electron microscopy. Subsequently my Director, Ray Bailey, gave me the encouragement and provided the impetus for me to embark on this present course of study.

Throughout this thesis the experimental results obtained by electron microscopy have, where possible, been related to biochemical and X-ray diffraction data kindly made available to me by Barbara Brodsky, Eric Eikenberry, Michael Flint, Gerry Gillard and Isabel Williams. Gary Thomas and Bob Fletcher supplied me with histogram plotting and population deconvolution programs and together provided me with oft-needed statistical advice. Doug Hopcroft has been responsible for the excellent maintenance of the electron microscopes and Ray Bennett printed the micrographs. I was assisted by many friends in typing the manuscript but it was June Tipoki who bore the lion's share of this chore.

Finally, and of greatest importance to me, my research colleague and supervisor, David Parry, not only instigated this project but displayed an un-ending enthusiasm for it. I am indebted to his necessary and continual encouragement throughout my writing-up.

To all of these people, and to many others whom I have personally acknowledged, I sincerely thank you for your support.

(v)

TABLE OF CONTENTS

ABSTRACT	iii
ACKNOWLEDGEMENTS	v
TABLE OF CONTENTS	vi
LIST OF TABLES	x
LIST OF FIGURES	xiii

CHAPTER 1: THE COLLAGENOUS COMPONENT OF CONNECTIVE TISSUES

1.1	INTRODUCTION	1
1.2	THE STRUCTURAL HIERARCHY OF COLLAGEN	
	1.2.1 The collagen molecule	2
	1.2.2 The collagen fibril	8
1.3	THE NATURE OF COLLAGENOUS TISSUES	
	1.3.1 The cellular components	11
	1.3.2 The fibrous components	14
1.4	SCOPE AND AIMS OF THIS THESIS	18
CHAPTER	2: MATERIALS AND METHODS	
2.1	INSTRUMENTAL METHOD	21
2.2	COLLECTION OF SPECIMENS	30
2.3	PREPARATIVE PROCEDURES	33
2.4	SECTIONING AND STAINING TECHNIQUES	34
2.5	PHOTOMICROGRAPHY AND MENSURATION METHODS	35
2.6	ANALYTICAL METHODS	37
CHAPTER	3: THE CORNEA	
3.1	INTRODUCTION	39
3.2	RESULTS AND DISCUSSION	47

Page

CHAPTER 4: EXPERIMENTAL OBSERVATIONS ON THE GROWTH AND

	DEVELOPMENT OF COLLAGEN FIBRILS	
4.1	INTRODUCTION	61
4.2	TENDONS AND LIGAMENTS	
	4.2.1 Avian Metatarsal Tendon	63
	4.2.2 Mammalian Tendons and Ligaments	70
4.3	SKINS	84
4.4	OTHER TISSUES	89
CHAPTER	5: COLLAGEN FIBRIL ASSEMBLY DISORDERS	
5.1	INTRODUCTION	93
5.2	FIBRILLAR MALFORMATIONS	
	5.2.1 Hereditable Disorders	96
	5.2.2 Naturally Occurring Malformations	102
5.3	CHANGING FIBRIL DIAMETER DISTRIBUTIONS	
	5.3.1 Induced Disorders	105
	5.3.2 Acquired Disorders	116
5.4	CONCLUSIONS	120
CHAPTER	6: INTERPRETATION AND SIGNIFICANCE OF COLLAGEN FIBRIL	
	DIAMETER DISTRIBUTION DATA	
6.1	CONSIDERATIONS OF THE LIMITATIONS IMPOSED BY ELECTRON	
	MICROSCOPY	123
6.2	ANALYSIS OF THE DIAMETER DISTRIBUTIONS THAT ARE SHARP	
	AND UNIMODAL	128
6.3	ANALYSIS OF DIAMETER DISTRIBUTIONS THAT ARE	
	HETEROGENEOUS	139
6.4	ANALYSIS OF BROAD DISTRIBUTIONS OF COLLAGEN FIBRIL	
	DIAMETER	141
6.5	CORRELATION BETWEEN ELECTRON MICROSCOPE AND X-RAY DATA	143
6.6	CONCLUSIONS	147
	(vii)	

CHAPTER 7: THE GROWTH AND DEVELOPMENT OF CONNECTIVE TISSUES

AND THE RELATIONSHIP BEIWEEN COLLAGEN FIBRIL DIAMETER

DISTRIBUT	TIONS AND MECHANICAL PROPERTIES	
7.1 INTRODUCT	TION	149
7.2 CORRELAT	IONS BETWEEN COLLAGEN FIBRIL DIAMETER	
DISTRIBUT	TIONS AND TISSUE ATTRIBUTES	
7.2.1 Foe	etal Development	152
7.2.2 Mod	des of Collagen Fibril Development for	
Alt	cricious and Precocious Animals	155
7.2.3 Pos	st-natal Development	157
7.2.4 For	m of the Collagen Fibril Diameter Distribution	
at	Maturity	159
7.2.5 For	m of the Collagen Fibril Diameter Distribution	
at	Senescence	164
7.2.6 Con	rrelation Between Mass-average Diameter and	
Ult	imate Tensile Strength	164
7.2.7 For	m of the Diameter Distribution and the	
Mec	chanical Properties of the Tissue	165
CHAPTER 8: THE RE	ELATIONSHIP BETWEEN GLYCOSAMINOGLYCAN	
COMPOSIT	ION AND COLLAGEN FIBRIL DIAMETERS: A POSSIBLE	
MECHANIS	1 FOR FIBRILLOGENESIS	
8.1 INTRODUCT	TION	170
8.2 DO GLYCOS	SAMINOGLYCANS MEDIATE CONTROL?	
8.2.1 Pre	evious Concepts	172
8.2.2 Obs	servations	175
8.2.3 Hyp	pothesis	184
8.3 CONCLUSIO	DNS	190
CHAPTER 9: SUMMAR	RY	195

(viii)

Page

APPENDICES

APPENDIX 1: GENETICALLY DISTINCT COLLAGEN TYPES AND THEIR	
DISTRIBUTION IN THE BODY TISSUES	200
APPENDIX 2: DIMENSIONS OF THE PARAMETERS OF PROPOSED	
COLLAGEN SUB-FIBRILLAR ASSEMBLIES	201
APPENDIX 3: SOURCE OF CORNEAS FOR COMPARATIVE STUDY AND	
STATE OF PRESERVATION PRIOR TO PREPARATION FOR ELECTRON	
MICROSCOPY	202
APPENDIX 4: PROCESSED MATERIAL SUPPLIED BY OTHER RESEARCH	
WORKERS	203
APPENDIX 5: BACTERIAL COLLAGENASE TREATED FLEXOR TENDONS	
FROM HORSE. EXPERIMENTAL PROTOCOL, CLINICAL AND	
POST-MORTEM OBSERVATIONS.	203a
REFERENCES	204

LIST OF TABLES

		Page
Table l.l	Components of connective tissues in mature animals.	16
Table 3.1	Mean diameters of populations of collagen fibrils	
	from the corneal stromal lamellae of the adult	
	vertebrates studied.	48
Table 3.2	Mean collagen fibril diameters of the corneal stroma	
	as cited in various works.	52
Table 3.3	Mean diameters of populations of collagen fibrils	
	from corneal stromal lamellae in developing frog,	
	rat, guinea pig, man and some neonatal mammals.	55
Table 3.4	Variation of mean collagen fibril diameter with depth	
	below the anterior surface of corneal stroma.	57
Table 3.5	Collagen fibrils of the vertebrate corneal stroma	
	having diameters which are not simple multiples of	
	~8 nm.	59
Table 4.1	Mean and mass-average diameters of collagen fibrils	
	in chick metatarsal tendons.	61
Table 4.2	Resolution of multimodal distributions of collagen	
	fibrils in chick metatarsal tendons.	6 6
Table 4.3	Modal centre-to-centre and mean surface separations of	
	collagen fibrils in foetal metatarsal tendons.	69
Table 4.4	Precocious-altricious classification of neonate	
	placental mammals.	73
Table 4.5	Mean and mass-average diameters of collagen fibril	
	diameter distributions of tendons and ligaments in	
	the developing sheep.	74
Table 4.6	Resolution of sub-populations of collagen fibril	
	diameters in sheep tendons and ligaments; Their	

(x)

.

comparison with the sharp unimodal distribution . observed.

79

80

85

86

90

92

- Table 4.7 Mean and mass-average diameters of collagen fibril diameter distributions of tendons from neonate to adult guinea pigs.
- Table 4.8 Mean and mass-average diameters of collagen fibril diameter distributions of flexor tendons in foetal to mature rats.
- Table 4.9 Resolution of bimodal distributions of collagen fibril diameters in rat tendons and their comparison to the sharp unimodal distributions observed.
- Table 4.10 Mean and mass-average diameters of collagen fibril diameter distributions in skin.
- Table 4.11 Mean and mass-average diameters of collagen fibril diameter distributions recorded from some miscellaneous tissues.
- Table 5.1 Collagen fibril diameter distribution data from horse superficial digital flexor tendon after treatment with bacterial collagenase. 113
- Table 5.2 Resolution of sub-populations of collagen fibrils in horse superficial digital flexor tendon treated with bacterial collagenase. 114
- Table 6.1 Diameters of collagen fibrils as determined byelectron microscope and X-ray diffraction studies.145
- Table 7.1 Form of the collagen fibril diameter distributionin foetal to senescent tissues.153
- Table 7.2 Mean and mass-average diameters of collagen fibrils in tendons, ligaments and skins in perinatal animals. 154

Table 7.3 Birth-mass of animals expressed as percentages of

(xi)

|--|

4

Table 7.4	Maximum mass-average diameter of collagen fibrils in	
	adult connective tissues.	169
Table 8.1	Glycosaminoglycan content and mass-average collagen	
	fibril diameter in skin as a function of age.	177
Table 8.2	Glycosaminoglycan content and mass-average collagen	
	fibril diameter in tendon as a function of age.	179

158

.

LIST OF FIGURES

		Page
Figure 1.1	Space-filling model of the collagen molecule.	3
Figure 1.2	Schematic representation of the procollagen molecule.	5
Figure 1.3	Schematic representation of collagen synthesis and	
	fibrillogenesis.	6
Figure 1.4	Electron micrograph of negatively stained collagen	
	fibrils with a diagram showing that the D-period can	
	be accounted for by a regular staggering of collagen	
	molecules.	10
Figure 1.5	Electron micrograph of transverse sections of the	
	collagen fibrils in foetal rat tail-tendon.	12
Figure 1.6	Electron micrograph of transverse sections of the	
	collagen fibrils in adult rat tail-tendon.	13
Figure 1.7	Electron micrographs of transverse sections of	
	elastic fibres at varying stages of development.	17
Figure 2.1	Cross-sectional drawing of a transmission electron	
	microscope column.	24
Figure 2.2	Simplified ray diagram of a transmission electron	
×	microscope.	25
Figure 2.3	Ray diagram illustrating the depth of field in an	
	electromagnetic lens.	29
Figure 3.1	Electron micrographs of transverse sections through	
	(a) the total thickness of the corneal stromata of	
	the snake and (b) a portion of the corneal stromata	
	of the magpie.	41
Figure 3.2	Electron micrographs of transverse and longitudinal	
	sections of sutural fibres in the corneal stromata of	
	the dogfish.	42

- Figure 3.3 Electron micrographs of transverse sections of the collagen fibrils from the corneal stroma of (a) bony fish (goldfish) and (b) cartilaginous fish (stingray). 50
- Figure 3.4 Electron micrographs of transverse sections of the collagen fibrils from the corneal stroma of (a) cartilaginous fish (dogfish), (b) amphibian (salamander), (c) reptile (snake), (d) bird (magpie) and (e) mammal (rabbit).

51

67

68

71

- Figure 4.1 Low magnification electron micrograph of the cellular elements and collagen fibrils in an 18 day foetal chick metatarsal tendon.
- Figure 4.2 Electron micrographs showing (a) the sharp unimodal distribution of collagen fibril diameters in 14 day foetal chick metatarsal tendon and (b) a collagen fibril diameter distribution which may be resolved into several distinct populations in 18 day foetal chick metatarsal tendon.
- Figure 4.3 Electron micrographs of bacterially-contaminated 15 day chick metatarsal tendon showing collagen fibrils falling into "close-arrays".
- Figure 4.4 Frequency and mass distributions of collagen fibril diameters in sheep flexor tendons. 76
- Figure 4.5 Frequency and mass distributions of collagen fibril diameters in sheep extensor tendons. 77
- Figure 4.6 Frequency and mass distributions of collagen fibril diameters in sheep suspensory ligaments. 78
- Figure 4.7 Frequency and mass distributions of collagen fibril diameters in guinea pig flexor tendons. 81
- Figure 4.8 Frequency and mass distributions of collagen fibril
 - (xiv)

diameters in guinea pig extensor tendons.

82

Figure 4.9 Frequency and mass distributions of collagen fibril diameters in guinea pig diaphragmatic tendons. 83 Figure 4.10 Frequency and mass distributions of collagen fibril diameters in rat forelimb flexor tendons. 87 Figure 4.11 Frequency and mass distributions of collagen fibril diameters in rat hindlimb flexor tendons. 88 Figure 5.1 Electron micrographs of transverse sections of the collagen fibrils in lamb skin; (a) and (c) are from an animal suffering from dermatosparaxis and (b) is from a control. 97 Figure 5.2 Electron micrograph of transverse sections of the collagen fibrils in dermatosparactic lamb tendon. 97 Figure 5.3 Electron micrographs of transverse sections of the collagen fibrils in (a) normal greyhound dermis (b) dysplastic greyhound-dermis and (c) papillary layer of dysplastic greyhound-dermis. 100 Figure 5.4 Electron micrographs of sections through the dysplastic greyhound-dermis showing (a) abnormal lysosomal activity in a dermal fibrocyte and details of (b) lamellar, (c) fibrinoid, and (d) "electron-dense" lysosomal inclusions. 101 Figure 5.5 Electron micrographs of transverse sections of collagen fibrils from (a) unstretched and (b) stretched Cuvierian tubules of the sea cucumber Holothuria forskali. 104 Figure 5.6 Frequency and mass distributions of collagen fibril diameters in normal rat skin and in healing

"longitudinal" and "transverse" wounds. 107

(xv)

- Figure 5.7 Frequency and mass distributions of collagen fibril diameters taken from the right (contralateral) superficial digital flexor tendons of horses whose left superficial flexor tendon had been treated with bacterial collagenase.
- Figure 5.8 Frequency and mass distributions of collagen fibril diameters from the superficial digital flexor tendons of horses at various times after treatment with bacterial collagenase.

111

112

129

- Figure 5.9 Electron micrographs of transverse sections of the collagen fibrils in (a) normal horse superficial digital flexor tendon, (b) 24 hours after bacterial collagenase treatment and (c) 4 weeks after bacterial collagenase treatment.
- Figure 5.10 Frequency and mass distributions of collagen fibril diameters in Dupuytren's contracture and nodules, and in normal palmar fascia.
- Figure 6.1 Electron micrographs of (a) overfocus and (b) underfocus Fresnel fringes in a holey plastic support film.
- Figure 6.2 (a) Histogram showing all data obtained from sharp unimodal collagen fibril diameter distributions having means in the range 14 - 44 nm. (b) Graph showing relationship between observed collagen fibril diameters and hypothetical sub-fibrillar units. 131
- Figure 6.3 Diagram showing the projected dimensions of a cylindrical fibril lying obliquely in a thin section. 134 Figure 6.4 Bar diagram showing the spread of recorded diameters from sharp unimodal distributions and the differences

(xvi)

in measurement made by two observers.

Figure 6.5 Electron microraph of transverse sections of collagen fibrils in the developing lamprey skin. Fibrils are of uniform diameter and appear to have electron translucent "cores". 140

137

- Figure 6.6 Frequency distributions of collagen fibril diameters in 13 day foetal chick metatarsal tendons as measured by three independent observers. 142
- Figure 7.1 Electron micrographs showing transverse sections of the collagen fibrils from the skins of lamprey, rat and trout. 160
- Figure 7.2 Frequency and mass distributions of collagen fibril diameters in skins from lamprey, rat and trout. 161
- Figure 7.3 Electron micrographs showing the lamellar arrangement of the collagen fibrils in lamprey skin. 163
- Figure 7.4 Graphs of (a) tensile strength and mass-average collagen fibril diameter versus age for rat skin and (b) tensile strength and mass-average collagen fibril diameter versus age for rat-tail tendon. 166 Figure 8.1 Graphs of tissue percentage contents of hyaluronic
 - acid, chondroitin sulphate and dermatan sulphate versus collagen fibril mass-average diameter. 183

(xvii)